

SUPPLEMENTARY MATERIAL TO

Performance of carbon-coated magnetic nanocomposite in methylene blue and arsenate treatment from aqueous solution

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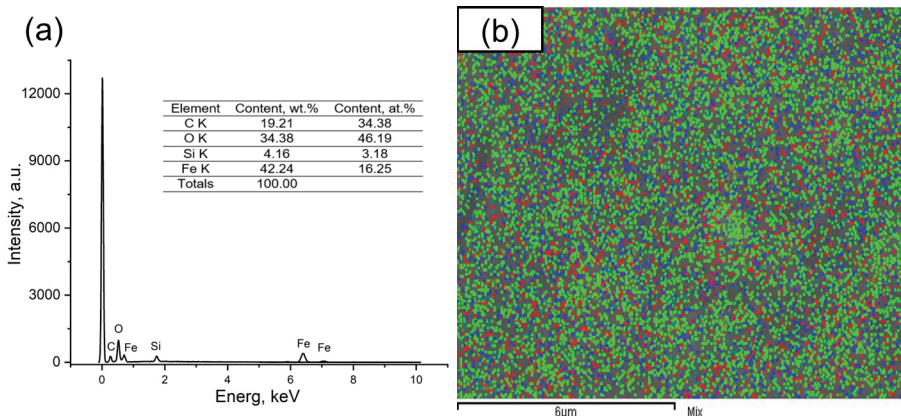


Fig. S-1. EDS analysis (a) and elemental map (b) of CMC.

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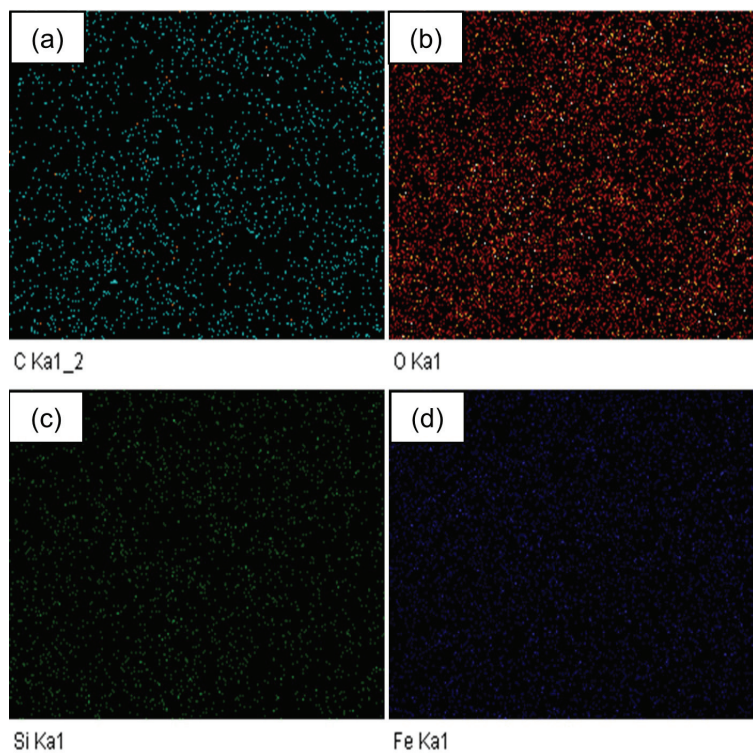


Fig. S-2. Elemental maps of C (a), O (b), Si (c) and Fe (d) of CMC.

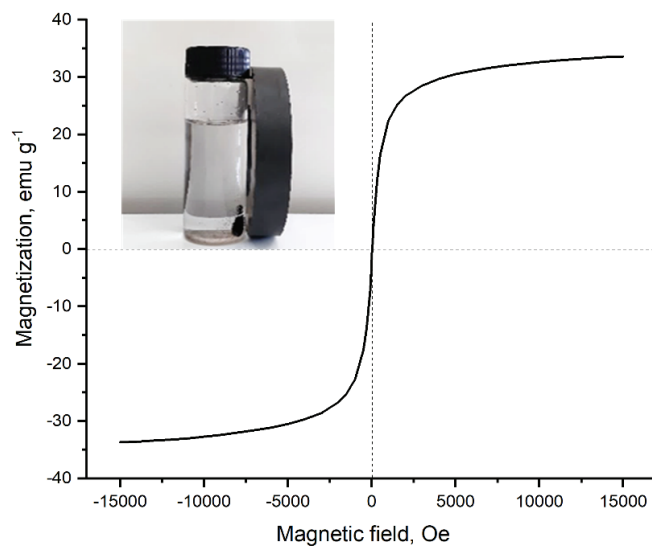


Fig. S-3. Magnetization curves and illustration of the magnetic separability of CMC.

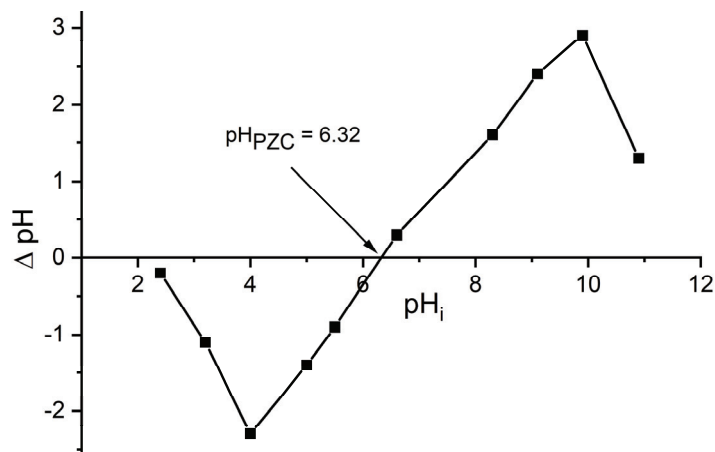


Fig. S-4. Plot of point of zero charge of CMC.

TABLE S-I. Different kinetic models, thermodynamic equations and adsorption isotherms

Model	Parameter	Equation
Adsorption kinetic models		
Pseudo first-order	$q_e / \text{mg g}^{-1}$ = equilibrium adsorption capacity	$q_t = q_e - q_e e^{-k_1 t}$ (1)
	$q_t / \text{mg g}^{-1}$ = adsorption capacity at time t	
	k_1 / min^{-1} = rate constant	
Pseudo second-order	$k_2 / \text{g mg}^{-1} \text{min}^{-1}$ = rate constant	$q_t = \frac{k^2 q_e^2 t}{1 + k^2 q_e t}$ (2)
Thermodynamic equations		
	$\Delta S^\circ / \text{J mol}^{-1}$ = entropy change	$\ln K_D = \frac{\Delta H^\circ}{RT} + \frac{\Delta S^\circ}{R}$ (3)
Van't Hoff equation	$\Delta H^\circ / \text{J mol}^{-1}$ = enthalpy change	
	$R / \text{J mol}^{-1} \text{K}^{-1} = 8.314$ (universal gas constant)	
	T / K = absolute temperature	
	$K_D / \text{L g}^{-1} = q_e / C_e$	
	thermodynamic equilibrium constant	
	$\Delta G^\circ / \text{J mol}^{-1}$ = Gibbs free energy change	$\square G^\circ = -RT \ln K_D$ (4)
Adsorption isotherms		
Langmuir	$q_m / \text{mg g}^{-1}$ = maximum monolayer adsorption capacity of the adsorbent	$\frac{C_e}{q_e} = \frac{1}{K_a q_m} + \frac{C_e}{q_m}$ (5)
	K_a = energy constant	
	R_L = separation factor which gives an idea about Langmuir isotherm	$R_L = \frac{1}{1 + K_a C_0}$ (6)
Freundlich	$K_F / \text{mg g}^{-1} \text{L}^{1/n} \text{mg}^{-1/n}$ = Freundlich constant n = intensity of adsorption, $n > 1$ indicates a favourable and heterogeneous adsorption	$\ln q_e = \ln K_F + \frac{1}{n} \ln C_e$ (7)

TABLE S-II. The comparison of the magnetization of CMC with various biochar

Precursors of magnetic biochar	Method	Magnetization, emu g^{-1}	Reference
Rice straw, $\text{Fe}(\text{NO}_3)_3$, KOH	Hydrothermal	33.7	This work
Coconut shells, FeCl_3	Pyrolysis, microwave	6.0	¹
Corn stalk, FeSO_4 , $\text{Na}_2\text{S}_2\text{O}_3$, NaOH	Hydrothermal	11.2	²
Corn stalk, FeSO_4 , $\text{Na}_2\text{S}_2\text{O}_3$, NaOH	Pyrolysis	20.4	²
Palm fiber, FeSO_4 , FeCl_3 , NH_3	Pyrolysis	19.4	³
Firwood, $\alpha\text{-FeOOH}$	Pyrolysis	20.8	⁴
Oleyl amine, FeCl_2 , FeCl_3 , NaOH	Hydrothermal	21.7	⁵
Rice husk, $\text{Fe}(\text{NO}_3)_3$, KMnO_4	Pyrolysis	27.5	⁶

TABLE S-III. The porous parameters of RS, BS, CMC samples

Sample	$S_{\text{BET}} / \text{m}^2 \text{g}^{-1}$	$V_{\text{T}} / \text{cm}^3 \text{g}^{-1}$	D_{p} / nm
RS	1.3	0.01	30.6
BS	6.6	0.04	33.0
CMC	171.4	0.15	6.0

TABLE S-IV. The comparison of the maximum adsorption capacity of MB and As(V) on CMC with various adsorbents.

Adsorbent	Capacity, mg g^{-1}		
	MB	As(V)	Ref.
CMC	110.63	2.31	This study
$\text{Fe}_2\text{O}_3\text{-ZrO}_2\text{/BC}$	38.1	1.01	⁷
M-MWCNTs	48.06	-	⁸
$\text{Fe}_3\text{O}_4\text{/MWCNT}$	74	-	⁹
$\text{Fe}_3\text{O}_4\text{@C NPs}$	117	-	¹⁰
HPB (hematite/biochar)	-	0.43	¹¹
Ch-Rs (chitosan/red scoria)	-	0.72	¹²
OBC (Canola straw-based biochar)	-	0.95	¹³
TB 800 (biochar from waste)	-	1.25	¹⁴
PAC-500 (magnetic biosorbents)	-	2.00	¹⁵
MC-O/NC-L-MG (magnetite/ microcellulose)	-	18.5	¹⁶
ChM (Chitosan-Magnetite Hydrogel)	-	66.9	¹⁷

REFERENCES

1. M. W. Yap, N. M. Mubarak, J. N. Sahu, E. C. Abdullah, *J. Ind. Eng. Chem.* **45** (2017) 287 (<https://doi.org/10.1016/j.jiec.2016.09.036>)
2. Y. Tu, Z. Peng, P. Xu, H. Lin, X. Wu, L. Yang, J. Huang, *Bioresources* **12** (2017) 1077 (<https://doi.org/10.15376/biores.12.1.1077-1089>)
3. X. Zhou, J. Zhou, Y. Liu, J. Guo, J. Ren, F. Zhou, *Fuel* **233** (2018) 469 (<https://doi.org/10.1016/j.fuel.2018.06.075>)
4. D. D. Sewu, H. N. Tran, G. Ohemeng-Boahen, S. H. Woo, *Sci. Total Environ.* **717** (2020) 137091 (<https://doi.org/10.1016/j.scitotenv.2020.137091>)
5. X. Bao, Z. Qiang, J.-H. Chang, W. Ben, J. Qu, *J. Environ. Sci.* **26** (2014) 962 ([https://doi.org/10.1016/S1001-0742\(13\)60485-4](https://doi.org/10.1016/S1001-0742(13)60485-4))

6. C. Sun, T. Chen, Q. Huang, J. Wang, S. Lu, J. Yan, *Environ. Sci. Pollut. Res. Int.* **26** (2019) 8902 (<https://doi.org/10.1007/s11356-019-04321-z>)
7. S. I. Siddiqui, S. A. Chaudhry, *J. Clean. Prod.* **223** (2019) 849 (<https://doi.org/10.1016/j.jclepro.2019.03.161>)
8. L. Ai, C. Zhang, F. Liao, Y. Wang, M. Li, L. Meng, J. Jiang, *J. Hazard. Mater.* **198** (2011) 282 (<https://doi.org/10.1016/j.jhazmat.2011.10.041>)
9. A. Suwattanamala, N. Bandis, K. Tedsree, C. Issro, *Mater. Today: Proc.* **4** (2017) 6567 (<https://doi.org/10.1016/j.matpr.2017.06.169>)
10. R. Wu, J.-H. Liu, L. Zhao, X. Zhang, J. Xie, B. Yu, X. Ma, S.-T. Yang, H. Wang, Y. Liu, *J. Environ. Chem. Eng.* **2** (2014) 907 (<https://doi.org/10.1016/j.jece.2014.02.005>)
11. S. Wang, B. Gao, A. R. Zimmerman, Y. Li, L. Ma, W. G. Harris, K. W. Migliaccio, *Bioresour. Technol.* **175** (2015) 391 (<https://doi.org/10.1016/j.biortech.2014.10.104>)
12. T. G. Asere, S. Mincke, J. De Clercq, K. Verbeken, D. A. Tessema, F. Fufa, C. V. Stevens, G. Du Laing, *Int. J. Environ. Res. Public Health* **14** (2017) 1 (<https://doi.org/10.3390/ijerph14080895>)
13. K. Zoroufchi Benis, J. Soltan, K. N. McPhedran, *Chem. Eng. J.* **423** (2021) 130061 (<https://doi.org/10.1016/j.cej.2021.130061>)
14. L. Verma, J. Singh, *J. Environ. Manage.* **248** (2019) 109235 (<https://doi.org/10.1016/j.jenvman.2019.07.006>)
15. L. Verma, M. A. Siddique, J. Singh, R. N. Bharagava, *J. Environ. Manage.* **250** (2019) 109452 (<https://doi.org/10.1016/j.jenvman.2019.109452>)
16. K. Taleb, J. Markovski, Z. Veličković, J. Rusmirović, M. Rančić, V. Pavlović, A. Marinković, *Arab. J. Chem.* **12** (2019) 4675 (<https://doi.org/10.1016/j.arabjc.2016.08.006>)
17. I. P. Verduzco-Navarro, E. Mendizabal, J. A. Rivera Mayorga, M. Renteria-Urquiza, A. Gonzalez-Alvarez, N. Rios-Donato, *Gels* **8** (2022) 1 (<https://doi.org/10.3390/gels8030186>).