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SUPPLEMENTARY MATERIAL TO Application of the redox system of *Nocardia corallina* B-276 in the enantioselective biotransformation of ketones and alcohols

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Fig. S-1. Reduction of ketone **1a** with *N. corallina* biomass, obtained at the final pH of the culture medium (5.36, 5.67 and 7.62).

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Fig. S-2. Biotransformation of **2a** with *N. corallina* biomass, obtained at the uncontrolled pH of the culture medium (4.74, 5.65 and 6.00).



Fig. S-3. Biotransformation of **3a** with *N. corallina* biomass obtained at the uncontrolled pH of the culture medium (5.80 and 7.30).





Fig. S-4. Biotransformation of **1b** with *N. corallina* biomass, final pH 7.62.



Fig. S5. Biotransformation of **2b** with *N. corallina* biomass obtained at the uncontrolled pH of the culture medium (5.99, 6.18, 6.28 and 6.80).



Fig. S-7. IR spectrum of 1-phenylpropan-1-ol (1b).

The ¹*H*- *and* ¹³*C*-*NMR spectra of* 1-*phenyl*-1-*propanone* (1*a*)

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 1.20 (3H, *t*, *J* = 7.3 Hz, CH₃), 2.96 (2H, *q*, *J* = 7.2 Hz, CH₂), 7.42 (2H, *t*, *J* = 7.7 Hz, CH), 7.51 (1H, *t*, *J* = 7.4 Hz, CH), 7.94 (2H, *dd*, *J* = 8.4 & 1.3 Hz, CH).

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¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 8.4 (C10), 31.7 (C8), 127.9/128.5 (C4, 3, 6 & 1), 132.9 (C2), 136.9 (C5), 200.7 (C7).



Fig. S-9. ¹³C-NMR spectrum of 1-phenyl-1-propanone (1a), in CDCl₃.

The ¹H- and ¹³C-NMR spectra of 1-phenylpropan-1-ol (1b)

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 0.77 (3H, *t*, *J* = 7.5 Hz, CH₃), 1.61 (2H, *ddt*, *J* = 2.1, 13.6 & 6.9 Hz, CH₂), 3.99 (1H, *s*, OH), 4.41 (1H, *t*, *J* = 6.7 Hz, CH), 7.13–7.19 (5H, *dt*, *J* = 12.8 & 6.9 Hz, CH).



Fig. S-10. ¹H-NMR spectrum of 1-phenylpropan-1-ol (1b), in CDCl₃.

¹³C-NMR (100 MHz, CDCl₃, δ / ppm): δ 10.2 (C10), 31.9 (C8), 75.7 (C7), 126.2 (C2), 127.3 (C6 & C4), 128.3 (C1 & C3) and 144.9 (C5) ppm.



Fig. S-11. ¹³C-NMR spectrum of 1-phenylpropan-1-ol (1b), in CDCl₃.

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The GC chromatograms of **1a** and **1b** with retention times of 4.39 and 5.63 min, respectively under conditions reported in the Experimental Section.



Fig. S-12. GC chromatograms of 1-phenyl-1-propanone (1a) and 1-phenylpropan-1-ol (1b).

The HPLC chromatogram of a mixture of 1a & rac-(1b), separately 1a and (S)-1b & (R)-1b with retention times of: (20.4, 11.4 and 9.6 min, respectively, column OB-H and 7.7, 14.9 and 12.7 min, respectively, column OD) with conditions reported in the Experimental Section.



Fig. S-13. HPLC chromatogram (OB-H column) of mixture of 1-phenyl-1-propanone (1a) & *rac*-1-phenylpropan-1-ol (1b), separately 1a and (S)-1b & (R)-1b.

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Fig. S-14. HPLC chromatogram (OD column) of a mixture of 1-phenyl-1-propanone (1a) & *rac*-1-phenylpropan-1-ol (1b), separately 1a and (S)-1b & (R)-1b.



Fig. S-16. IR spectrum of 1-phenyl-1,2-ethanediol (2b).

The ¹H- and ¹³C-NMR spectra of 2-hydroxy-1-phenylethanone (2a)

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 4.9 (2H, *m*, CH₂), 5.4 (1H, *s*, OH), 7.5 (2H, *dd*, *J* = 11.9 & 3.9 Hz, CH), 7.63 (1H, *t*, *J* = 7.7 Hz, CH), 7.93 (2H, *dd*, *J* = 8.3 & 1.1 Hz, CH).





Fig. S-17. ¹H-NMR spectrum of 2-hydroxy-1-phenylethanone (2a) in CDCl₃.







The ¹H- and ¹³C-NMR spectra of 1-phenyl-1,2-ethanediol (2b)

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 2.3 (1H, *s*, OH), 3.2 (1H, *s*, OH), 3.49–3.78 (2H, *m*, CH₂) 4.8 (1H, *dd*, *J* = 8.4 & 3.3 Hz, CH), 7.09–7.48 (5H, *m*, CH).



Fig. S-19. ¹H-NMR spectrum of 1-phenyl-1,2-ethanediol (2b) in CDCl₃.

¹³C-NMR (100 MHz, CDCl₃ δ / ppm,): 67.95/68.04 (C8), 74.7 (C7), 126.07 (C4 & C6), 127.9 & 128.5 (C1, 2 & C3), 140.5 (C5).



Fig. S-20. ¹³C-NMR spectrum of 1-phenyl-1,2-ethanediol (2b), in CDCl₃.

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The GC chromatograms of **2a** and **2b** with retention times of 4.90 and 5.79 min, respectively, with conditions reported in the Experimental Section.

Fig. S-21. GC chromatograms of 2-hydroxy-1-phenylethanone (2a) and 1-phenyl-1,2-ethanediol (2b) and a mixture of 2a and *rac*-2b.

The HPLC chromatograms of mixtures of 2a, (S)-2b and (R)-2b with retention times of 17.12, 11.38 and 9.04 min, respectively, with conditions reported in the Experimental Section.



Fig. S-22. HPLC chromatogram of: I) mixture of 2-hydroxy-1-phenylethanone (2a) and *rac*-1-phenyl-1,2-ethanediol (*rac*-2b); II) 2-hydroxy-1-phenylethanone (2a);
III) *rac*-1- phenyl-1,2-ethanediol (*rac*-2b); IV) (*R*)-1-phenyl-1,2-ethanediol (*R*-2b);
V) (*S*)-1-phenyl-1,2-ethanediol (*S*2b)

The HPLC chromatograms of mixtures of 2a, (S)-2b and (R)-2b with retention times of 28.9, 18.9 and 15.1 min, respectively, under the conditions reported in the Experimental Section.

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Fig. S-23. HPLC chromatogram of: I) 2-hydroxy-1-phenylethanone (**2a**); II) *rac*-1-phenyl-1,2-ethanediol (*rac*-**2b**); III) (*R*)-1-phenyl-1,2-ethanediol (*R*-**2b**); IV) (*S*)-1-phenyl-1,2-ethanediol ((*S*)-**2b**).

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Fig. S-25. IR spectrum of methyl (2-chlorophenyl)(hydroxy)acetate (3b).

The ¹H- and ¹³C-NMR spectra of methyl (2-chlorophenyl)(oxo)acetate (3a)

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 3.96 (3H, *s*), 7.43 (2H, *m*), 7.53 (1H, *ddd*, *J* =8.0, 7.4 & 1.7 Hz), 7.77 (1H, *dd*, *J* =7.7 & 1.7 Hz).

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Fig. S-26. ¹H-NMR spectrum of methyl (2-chlorophenyl)(oxo)acetate (3a), in CDCl₃



Fig. S-27. ¹³C-NMR spectrum of methyl (2-chlorophenyl)(oxo)acetate (3a), in CDCl₃.

¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 53.2 (C9), 127.2 (C4), 130.5 (C2), 131.6 (C5), 133.2 (C6), 133.9 (C1), 134.3 (C3), 163.4 (C8), 186.2 (C7).

The ¹*H*- *and* ¹³*C*-*NMR spectra of methyl (2-chlorophenyl)(hydroxy)acetate (3b)*

¹H-NMR (600 MHz, CDCl₃, δ / ppm): 3.6 (1H, *d*, *J* = 5.1 Hz, OH), 3.8 (3H, *s*, CH₃), 5.6 (1H, *d*, *J* = 5.1 Hz, CH) 7.3 (2H, *m*), 7.4 (2H, *m*).

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Fig. S-28. 1H-NMR spectrum of methyl (2-chlorophenyl)(hydroxy)acetate (3b), in CDCl₃.

¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 53.2 (C9), 70.3 (C7), 127.1 (C4), 128.8 (C2), 129.7(C5), 129.9 (C3), 133.41 (C1), 135.9 (C6), 173.7 (C8).



Fig. S-29. ¹³C-NMR spectrum of methyl (2-chlorophenyl)(hydroxy)acetate (3b), in CDCl₃.

The GC chromatograms of 3a and 3b with retention times of 4.6 and 6.1 min, respectively, under conditions reported in the Experimental Section.



Fig. S-30. GC chromatograms of methyl (2-chlorophenyl)(oxo)acetate (**3a**) and methyl (2-chlorophenyl)(hydroxy)acetate (**3b**).

The HPLC chromatogram of the mixture of 3a, (S)-3b and (R)-3b with retention times of 7.0, 8.9 and 10.2 min, respectively, under conditions reported in the Experimental Section.



Fig. S-31. HPLC chromatogram of: I) mixture of methyl (2-chlorophenyl)(oxo)acetate (**3a**) and *rac*-methyl (2-chlorophenyl)(hydroxy)acetate (*rac*-**3b**); II) methyl (2-chlorophenyl)(oxo)acetate (**3a**); III) *rac*-methyl (2-chlorophenyl)(hydroxy)acetate (*rac*-**3b**) and IV (*R*)-methyl (2-chlorophenyl)(hydroxy)acetate (*R*-**3b**).

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Fig. S-32. HPLC chromatogram of Fig. 2. Reduction of ketones **1a** with *N. corallina* biomass, final pH (5.36, 5.67 and 7.62).

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Fig. S-33. HPLC chromatogram of Fig. 3. Biotransformation of **1a** with *N. corallina* biomass, final pH 5.36.

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Fig. S-34. HPLC chromatogram of Fig. 4. Biotransformation of **2a** with *N. corallina* biomass, for different final pH values and times.



Fig. S-35. HPLC chromatogram of Fig. 5. Biotransformation of **3a** with *N. corallina* biomass, final pH 5.8 and 7.3.

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Fig. S-36. HPLC chromatogram of Fig. 6. Biotransformation of **1b** with *N. corallina* biomass, final pH 7.62.



Fig. S-37. HPLC chromatogram of Fig. 7. Biotransformation of **2b** with *N. corallina* biomass, at different final pH values and times.

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Fig. S-38. CG chromatogram of Fig. 8. Biotransformation of **2b** with *N. corallina* biomass, final pH 5.9.

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Fig. S-39. HPLC chromatogram of Fig. 8. Biotransformation of **2b** with *N. corallina* biomass, final pH 5.9.

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Fig. S-40. HPLC chromatogram of Fig. 9. Biotransformation of **3b** with *N. corallina* biomass, final pH 5.9.



Fig. S-41. HPLC chromatogram of Fig. 9. Biotransformation of **3b** with *N. corallina* biomass, final pH 7.3.